

SHORT COMMUNICATION

The effects of hypoxic bradycardia and extracellular $\text{HCO}_3^-/\text{CO}_2$ on hypoxic performance in the eel heart

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ABSTRACT

During hypoxia, fishes exhibit a characteristic hypoxic bradycardia, the functional significance of which remains debated. Here, we investigated the hypothesis that hypoxic bradycardia primarily safeguards cardiac performance. In preparations from the European eel (*Anguilla anguilla*), a decrease in stimulation frequency from 40 to 15 beats min^{-1} , which replicates hypoxic bradycardia *in vivo*, vastly improved cardiac performance during hypoxia *in vitro*. As eels display dramatic shifts in extracellular $\text{HCO}_3^-/\text{CO}_2$, we further investigated the effect this has upon hypoxic cardiac performance. Elevations from 10 mmol l^{-1} $\text{HCO}_3^-/1\%$ CO_2 to 40 mmol l^{-1} $\text{HCO}_3^-/4\%$ CO_2 had few effects on performance; however, further, but still physiologically relevant, increases to 70 mmol l^{-1} $\text{HCO}_3^-/7\%$ CO_2 compromised hypoxia tolerance. We revealed a four-way interaction between $\text{HCO}_3^-/\text{CO}_2$, contraction frequency, hypoxia and performance over time, whereby the benefit of hypoxic bradycardia was most prolonged at 10 mmol l^{-1} $\text{HCO}_3^-/1\%$ CO_2 . Together, our data suggest that hypoxic bradycardia greatly benefits cardiac performance, but its significance may be context specific.

KEY WORDS: Heart rate, Myocardial hypoxia, Contractility, *Anguilla anguilla*

INTRODUCTION

Fishes exhibit a characteristic slowing of the heart during hypoxia. This enigmatic hypoxic bradycardia is primarily mediated by increased vagal tone and hence reflects a regulated physiological response, but its adaptive significance, albeit extensively debated, remains elusive (Randall, 1982; Perry and Desforges, 2006; Farrell, 2007). Recent studies in eels (Iversen et al., 2010) and other teleosts (Perry and Desforges, 2006; McKenzie et al., 2009) demonstrate that pharmacological or surgical elimination of hypoxic bradycardia does not affect branchial oxygen uptake, suggesting that the potential benefit of the lower heart rate lies elsewhere in the cardiorespiratory system. Intriguingly, it has been suggested that hypoxic bradycardia represents a strategy to protect the heart itself (Farrell, 2007). This hypothesis is partly founded on the negative force–frequency effect of fish hearts, which describes the greater force generation at low heart rates due to the greater potential to reduce diastolic calcium levels and thus increase calcium transients (reviewed in Shiels et al., 2002). This may be particularly relevant during hypoxia and reoxygenation, when myocardial relaxation becomes impaired (e.g. Stecyk et al., 2011; Joyce et al., 2015). By way of analogy, it was recently demonstrated that simulated

cold-induced bradycardia ‘protects’ calcium transients at low temperature in bluefin tuna cardiomyocytes (Shiels et al., 2015).

Eels (*Anguilla* spp.) are particularly tolerant of hypoxia (Wood and Johansen, 1973; Iversen et al., 2010), at least in part because of an exceptional hypoxia tolerance of the heart (Gesser et al., 1982; Davie et al., 1992; Hartmund and Gesser, 1996). Environmental hypoxia is typically associated with other stressors, including hypercapnia, but how such extrinsic factors may modulate myocardial hypoxia tolerance is not well understood. Upon adaptation to freshwater (Farrell and Lutz, 1975) or exposure to chronic hypercapnia (McKenzie et al., 2003), European eels (*A. anguilla*) vastly elevate plasma bicarbonate concentrations ($[\text{HCO}_3^-]$) at the expense of chloride (Cl^-). During severe hypercapnia, $[\text{HCO}_3^-]$ may exceed 70 mmol l^{-1} , which is unparalleled amongst teleosts and facilitates effective pH regulation (McKenzie et al., 2003). The concomitant rise in $\text{HCO}_3^-/\text{CO}_2$ (at constant pH) enhances myocardial hypoxia tolerance in carp (*Cyprinus carpio*), and to a lesser extent rainbow trout (*Oncorhynchus mykiss*) (Gesser, 1977), although we recently failed to report such an effect in the air-breathing fish *Pangasianodon hypophthalmus* (Joyce et al., 2015). It is nevertheless possible that, as in carp, compensatory acid–base changes may confer an additional benefit to the eel heart during hypoxia.

Here, we investigate whether physiologically relevant changes in $\text{HCO}_3^-/\text{CO}_2$ and contraction frequency affect hypoxia tolerance in European eel cardiac preparations. We hypothesise that both elevated $\text{HCO}_3^-/\text{CO}_2$ and bradycardia confer cardiac protection during oxygen deprivation.

MATERIALS AND METHODS

Experimental animals

European eels (*Anguilla anguilla* [Linnaeus 1758]; 380–1000 g) of undetermined sex were obtained from a local eel farm (Lyksvad fish farm, Denmark) and maintained at Aarhus University in large aquaria (18°C) where they were fed commercial fish food twice a week.

Myocardial preparations

Eels were anaesthetised in an aerated benzocaine solution (0.2 g l^{-1}) until gill ventilation ceased, before the heart was rapidly excised and placed in ice-cold 10 mmol l^{-1} HCO_3^- Ringer’s solution (full composition below). Strips of myocardium were prepared and tied at each end with surgical silk, so one end could be secured to a metal rod attached to a force transducer, whilst the other end was anchored to one of two platinum electrodes in an organ bath containing 50 ml of Ringer’s solution at 18°C. The two platinum electrodes were connected to Grass stimulators (SD9), which provided 5 ms pulses at 0.66 Hz (40 beats min^{-1}) with a voltage adjusted to twice the minimum required to invoke full contraction. Preparations were then stretched with a micrometer screw to attain maximum force.

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Experimental protocol

Three different Ringer's solutions and gas mixtures were employed. In the first, 10 mmol l⁻¹ HCO₃⁻ Ringer's solution [NaCl (120 mmol l⁻¹), NaHCO₃ (10), KCl (4), NaH₂PO₄ (1), CaCl (3.5) and glucose (5)] was bubbled with 1% CO₂, 49% N₂ and 50% O₂. In the two other conditions, to elevate HCO₃⁻ whilst maintaining Na⁺ constant (131 mmol l⁻¹), NaCl was exchanged for equimolar NaHCO₃. NaHCO₃ was increased to 40 or 70 mmol l⁻¹, whilst NaCl was reduced to 90 or 60 mmol l⁻¹, respectively. In parallel, CO₂ was elevated to 4 and 7% at the expense of N₂, thus maintaining a pH of 7.65. This represents the range of HCO₃⁻/CO₂ reported in eels by McKenzie et al. (2003). CaCl was also elevated to 4 and 4.5 mmol l⁻¹ to mitigate for the effects of CaHCO₃ formation (Schaefer, 1974) and further mimic the conditions reported by McKenzie et al. (2003) during hypercapnia.

Four strips were run in parallel at the same HCO₃⁻/CO₂ level. After at least an hour of stabilization, two strips were rendered severely hypoxic by exchanging O₂ for N₂ for 30 min. During this period, the stimulation frequency of one normoxic and one hypoxic strip was decreased to 0.25 Hz (15 beats min⁻¹) to resemble the hypoxic bradycardia in eels (Iversen et al., 2010). Preparations were then re-oxygenated for 10 min at 0.66 Hz. The mass and length of the strips were measured upon completion of the experiment so that force could be calculated relative to cross-sectional area (e.g. Gesser, 1977). Preparations were between 5 and 10 mm in length with a mean (±s.e.m.) cross-sectional area of 1.40±0.14 mm².

Analyses

To investigate contractile force and diastolic dynamics, twitch force and the rate of 50% relaxation were recorded 10 min before hypoxia and at 5 min intervals thereafter. An additional recording was taken 2.5 min after reoxygenation to increase the temporal resolution during this critical period.

Statistical analysis was carried out in R and Prism (GraphPad). A one-way ANOVA was carried out on the absolute values of twitch force and rate of 50% relaxation at the start of the different experimental protocols. As there were no initial differences in force

($P=0.33$) or rate of 50% relaxation ($P=0.27$) between conditions, all values were normalised to those measured 10 min before hypoxia. The effects of time, hypoxia exposure, HCO₃⁻/CO₂ and stimulation frequency on relative changes in force and rate of 50% relaxation were analysed using a linear mixed effects model with hypoxia, HCO₃⁻/CO₂ and stimulation frequency treated as factors. All values are presented as means±s.e.m.

RESULTS AND DISCUSSION

A representative trace of contractile strength at 10 mmol l⁻¹ HCO₃⁻ and 1% CO₂ is depicted in Fig. 1. The linear mixed effects model revealed that time ($P<0.001$), hypoxia ($P<0.001$), stimulation frequency ($P<0.001$) and HCO₃⁻/CO₂ ($P=0.003$) all exerted independent effects on the relative change in twitch force. Further, hypoxia reduced the rate of relaxation, which persisted during re-oxygenation ($P<0.001$; Fig. 2).

There was a significant interaction between stimulation frequency and hypoxia ($P<0.001$; Fig. 2), whereby physiologically relevant decreases in stimulation frequency, to mimic the hypoxic bradycardia measured *in vivo* (Iversen et al., 2010), vastly improved hypoxia tolerance of the eel myocardium. This strongly supports Farrell's (2007) hypothesis that hypoxic bradycardia foremost protects cardiac performance. Indeed, at the frequencies investigated, there appeared to be little force–frequency dependency during normoxia, although this was accentuated at elevated HCO₃⁻/CO₂. In other words, severe hypoxia intensified the negative force–frequency effect. The benefit of lower heart rates during hypoxia is reminiscent of that previously characterised in fish during acute and chronic temperature changes (Shiels et al., 2002, 2015).

At low contraction frequencies, cardiomyocytes are better able to reduce diastolic calcium levels and thus increase the activator calcium transient (Harwood et al., 2000; Shiels et al., 2002). As relaxation becomes particularly impaired during hypoxia (Stecyk et al., 2011; Joyce et al., 2015), the greater inter-contraction interval will benefit calcium exudation during diastole. Further, in the present study we revealed greater rates of relaxation during hypoxia

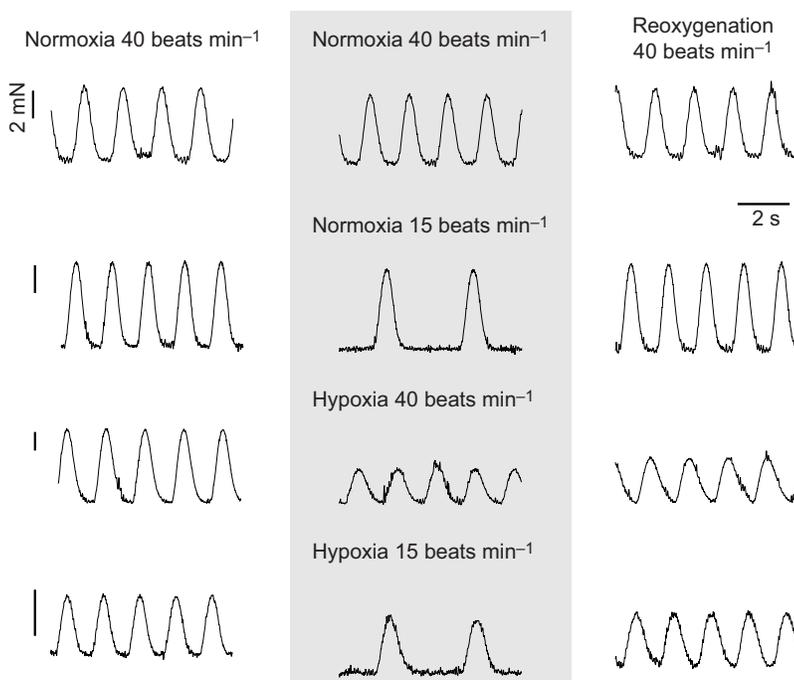


Fig. 1. Original and representative traces depicting the effect of hypoxia (30 min) and reoxygenation (5 min) and changes in stimulation frequency in eel (*Anguilla anguilla*) cardiac preparations (10 mmol l⁻¹ HCO₃⁻/1% CO₂; pH 7.65).

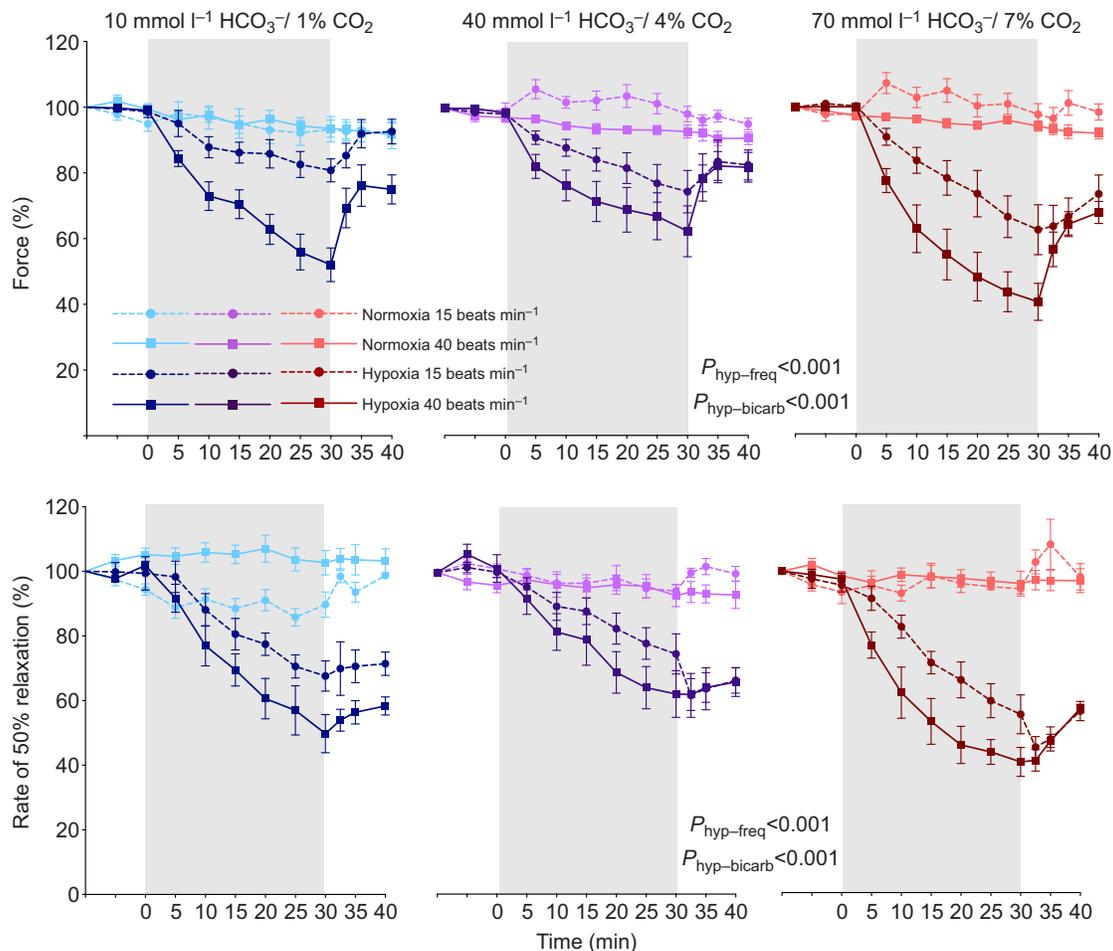


Fig. 2. The effect of stimulation frequency and variable $\text{HCO}_3^-/\text{CO}_2$ (pH 7.65) on twitch force and rate of 50% relaxation in eel (*Anguilla anguilla*) cardiac preparations during hypoxia and reoxygenation. The shaded area demarcates when preparations were made hypoxic and/or stimulation frequency was changed (see key). P -values represent the interaction between hypoxia (hyp) treatment and stimulation frequency (freq) or $\text{HCO}_3^-/\text{CO}_2$ (bicarb) (linear mixed effects model). $n=7$ in all cases except for $10 \text{ mmol l}^{-1} \text{ HCO}_3^-/1\% \text{ CO}_2$, normoxia $15 \text{ beats min}^{-1}$, in which case $n=6$.

at lower stimulation frequencies (Fig. 2). In the *in vivo* situation this is likely to reduce end-diastolic volume and thus maintain large stroke volumes during oxygen deprivation.

The hypoxic bradycardia may also benefit other aspects of cardiac calcium handling and excitation–contraction coupling. For example, low heart rates afford prolonged action potentials (Harwood et al., 2000; Shiels et al., 2015), which may increase the activator calcium influx during hypoxia and compensate for lowered calcium sensitivity by the myofilaments as inorganic phosphates rise (Driedzic and Gesser, 1994; Jensen and Gesser, 1999).

An additional value of hypoxic bradycardia may reside in myocardial energetics (Farrell, 2007). By estimating cardiac work from the product of contraction frequency and force, our data reveal that bradycardia may reduce myocardial energy demand by up to 50%, despite the increased twitch force. Indeed, because fish hearts display a characteristic ability to increase stroke volume at low heart rates (‘intrinsic autoregulation of cardiac output’ *sensu* Altimiras and Axelsson, 2004), the hypoxic bradycardia does not necessarily decrease cardiac output, although cardiac output of eels does decrease in very severe hypoxia (Iversen et al., 2010). ‘Saving’ energy on excitation–contraction coupling has the dual benefit of conserving cardiac energy stores and minimising the accumulation of lactic acid.

Further, the prolonged diastolic period may favour the removal of inorganic phosphates by glycolysis and prolong the time for energy restoration at ATPase sites.

Because eels exhibit large *in vivo* elevations in $[\text{HCO}_3^-]$ to above 70 mmol l^{-1} (McKenzie et al., 2003), i.e. more than twice that investigated in other species (Gesser, 1977; Joyce et al., 2015), we reasoned that any potential benefit of elevated $\text{HCO}_3^-/\text{CO}_2$ would be maximally revealed in the eel heart. However, in contrast to our initial hypothesis, increasing $\text{HCO}_3^-/\text{CO}_2$ to 70 mmol l^{-1} and 7%, respectively, reduced both force generation and the rate of relaxation during hypoxia ($P<0.001$; Fig. 2). Preparations at high $\text{HCO}_3^-/\text{CO}_2$ also performed particularly poorly during re-oxygenation.

When comparing with other fish species, such as the common carp in which protective effects of elevated $\text{HCO}_3^-/\text{CO}_2$ appear most marked (Gesser, 1977), it is important to note that eel preparations were considerably more hypoxia tolerant than carp (20% reduction in force versus 65% reduction in force during similar experimental conditions) at similar and low $\text{HCO}_3^-/\text{CO}_2$ concentrations ($10 \text{ mmol l}^{-1}/1\%$ and $6.25 \text{ mmol l}^{-1}/1\%$, respectively). However, at higher $\text{HCO}_3^-/\text{CO}_2$ ($40 \text{ mmol l}^{-1}/4\%$ and $25 \text{ mmol l}^{-1}/4\%$), eels and carp compared much more favourably (20–25% reduction in force in both cases). This suggests that eel myocardium cannot benefit from the additional

increases in bicarbonate that the carp heart can exploit. Indeed, it is not surprising that the hypoxia tolerance of the eel myocardium could not be improved given that, at low heart rate, hypoxic preparations were remarkably similar to normoxic controls. The detrimental effect of 70 mmol l⁻¹ HCO₃⁻/7% CO₂ was unexpected, but may be rationalised based on the acute nature of the experiment. It is possible that the preparations may have been compromised by the low [Cl⁻] needed to compensate for elevated [HCO₃⁻]. In the mammalian heart, chloride currents appear important in the recovery from ischaemia–reperfusion events (Petrich et al., 1996; Uramoto et al., 2012), but this remains to be investigated in piscine hearts. Given that elevated plasma [HCO₃⁻] levels in response to severe hypercapnia did not affect *in vivo* hypoxia tolerance in eels (McKenzie et al., 2003), the potential detriment reported in our study may be compensated for during acclimation.

HCO₃⁻/CO₂ affected the importance of stimulation frequency during hypoxia over time (i.e. there was a four-way interaction between time, HCO₃⁻/CO₂, hypoxia treatment and stimulation frequency, *P*<0.001). The beneficial effect of bradycardia was clearly discernible within the first 10 min of hypoxia, whilst twitch force declined linearly thereafter. To further explore this relationship, linear regressions were performed on the fall of twitch force between 10 and 30 min of hypoxia exposure at different HCO₃⁻/CO₂ concentrations. At elevated HCO₃⁻/CO₂ (40 or 70 mmol l⁻¹ HCO₃⁻ and 4 or 7% CO₂), the slopes of 15 and 40 beats min⁻¹ preparations were indistinguishable (*P*=0.95 in both cases). However, at 10 mmol l⁻¹ HCO₃⁻ and 1% CO₂, preparations paced at 40 beats min⁻¹ had a significantly steeper slope than those at 15 beats min⁻¹ (*P*=0.04). Thus, lower HCO₃⁻/CO₂ prolonged the benefit of hypoxic bradycardia. This difference was further evident during reoxygenation, when only bradycardic 10 mmol l⁻¹ HCO₃⁻/1% CO₂ preparations fully recovered twitch force. The preparations that did not fully recover force may have incurred permanent damage during the hypoxic insult. Further, at elevated HCO₃⁻/CO₂, the rate of relaxation during reoxygenation in preparations stimulated at different frequencies entirely converged, whereas at 10 mmol l⁻¹ HCO₃⁻ and 1% CO₂, bradycardic preparations maintained faster relaxation rates during recovery.

In conclusion, our data support the notion that the hypoxic bradycardia benefits myocardial performance, but also emphasise that the functional importance of this benefit depends on other environmental and physiological factors.

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Competing interests

The authors declare no competing or financial interests.

Author contributions

W.J. conceived and designed the experiments, M.S. and W.J. performed the experiments and analysed the data, and all authors contributed to writing the paper.

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References

- Altimiras, J. and Axelsson, M. (2004). Intrinsic autoregulation of cardiac output in rainbow trout (*Oncorhynchus mykiss*) at different heart rates. *J. Exp. Biol.* **207**, 195–201.
- Davie, P. S., Farrell, A. P. and Franklin, C. E. (1992). Cardiac performance of an isolated eel heart: effects of hypoxia and responses to coronary artery perfusion. *J. Exp. Zool.* **262**, 113–121.
- Driedzic, W. R. and Gesser, H. (1994). Energy metabolism and contractility in ectothermic vertebrate hearts – hypoxia, acidosis and low temperature. *Physiol. Rev.* **74**, 221–258.
- Farrell, A. P. (2007). Tribute to P. L. Lutz: a message from the heart – why hypoxic bradycardia in fishes? *J. Exp. Biol.* **210**, 1715–1725.
- Farrell, A. P. and Lutz, P. L. (1975). Apparent anion imbalance in the fresh water adapted eel. *J. Comp. Physiol.* **102**, 159–166.
- Gesser, H. (1977). Effects of hypoxia and reoxygenation on force development in myocardia of carp and rainbow trout: protective effects of CO₂/HCO₃⁻. *J. Exp. Biol.* **69**, 199–206.
- Gesser, H., Andresen, P., Brams, P. and Sund-Laursen, J. (1982). Inotropic effects of adrenaline on the anoxic or hypercapnic myocardium of rainbow trout and eel. *J. Comp. Physiol. B* **147**, 123–128.
- Hartmund, T. and Gesser, H. (1996). Cardiac force and high-energy phosphates under metabolic inhibition in four ectothermic vertebrates. *Am. J. Physiol.* **271**, R946–R954.
- Harwood, C. L., Howarth, F. C., Altringham, J. D. and White, E. (2000). Rate-dependent changes in cell shortening, intracellular Ca²⁺ levels and membrane potential in single, isolated rainbow trout (*Oncorhynchus mykiss*) ventricular myocytes. *J. Exp. Biol.* **203**, 493–504.
- Iversen, N. K., McKenzie, D. J., Malte, H. and Wang, T. (2010). Reflex bradycardia does not influence oxygen consumption during hypoxia in the European eel (*Anguilla anguilla*). *J. Comp. Physiol. B* **180**, 495–502.
- Jensen, M. A. and Gesser, H. (1999). Influence of inorganic phosphate and energy state on force in skinned cardiac muscle from freshwater turtle and rainbow trout. *J. Comp. Physiol. B* **169**, 439–444.
- Joyce, W., Gesser, H., Bayley, M. and Wang, T. (2015). Anoxia and acidosis tolerance of the heart in an air-breathing fish (*Pangasianodon hypophthalmus*). *Physiol. Biochem. Zool.* **88**, 648–659.
- McKenzie, D. J., Piccolella, M., Dalla Valle, A. Z., Taylor, E. W., Bolis, C. L. and Steffensen, J. F. (2003). Tolerance of chronic hypercapnia by the European eel *Anguilla anguilla*. *J. Exp. Biol.* **206**, 1717–1726.
- McKenzie, D. J., Skov, P. V., Taylor, E. W. T., Wang, T. and Steffensen, J. F. (2009). Abolition of reflex bradycardia by cardiac vagotomy has no effect on the regulation of oxygen uptake by Atlantic cod in progressive hypoxia. *Comp. Biochem. Physiol.* **153**, 332–338.
- Perry, S. F. and Desforges, P. R. (2006). Does bradycardia or hypertension enhance gas transfer in rainbow trout (*Oncorhynchus mykiss*)? *Comp. Biochem. Physiol. A* **144**, 163–172.
- Petrich, E. R., Zumino, A. P. and Schanne, O. F. (1996). Early action potential shortening in hypoxic hearts: Role of chloride current(s) mediated by catecholamine release. *J. Mol. Cell Cardiol.* **28**, 279–290.
- Randall, D. (1982). The control of respiration and circulation in fish during exercise and hypoxia. *J. Exp. Biol.* **100**, 275–288.
- Schaer, H. (1974). Decrease in ionized calcium by bicarbonate in physiological solutions. *Pflügers Arch.* **347**, 249–254.
- Shiels, H. A., Vornanen, M. and Farrell, A. P. (2002). The force–frequency relationship in fish hearts—a review. *Comp. Biochem. Physiol. A* **132**, 811–826.
- Shiels, H. A., Galli, G. L. J. and Block, B. A. (2015). Cardiac function in an endothermic fish: cellular mechanisms for overcoming acute thermal challenges during diving. *Proc. R. Soc. B Biol. Sci.* **282**, 20141989.
- Stecyk, J. A. W., Larsen, B. C. and Nilsson, G. E., (2011). Intrinsic contractile properties of the crucian carp (*Carassius carassius*) heart during anoxic and acidotic stress. *Am. J. Physiol.* **301**, R1132–R1142.
- Uramoto, H., Okada, T. and Okada, Y. (2012). Protective role of cardiac CFTR activation upon early reperfusion against myocardial infarction. *Cell Physiol. Biochem.* **30**, 1023–1038.
- Wood, S. C. and Johansen, K. (1973). Blood oxygen transport and acid-base balance in eels during hypoxia. *Am. J. Physiol.* **225**, 849–851.