RESEARCH ARTICLE

Environment, behavior and physiology: do birds use barometric pressure to predict storms?

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SUMMARY

Severe storms can pose a grave challenge to the temperature and energy homeostasis of small endothermic vertebrates. Storms are accompanied by lower temperatures and wind, increasing metabolic expenditure, and can inhibit foraging, thereby limiting energy intake. To avoid these potential problems, most endotherms have mechanisms for offsetting the energetic risks posed by storms. One possibility is to use cues to predict oncoming storms and to alter physiology and behavior in ways that make survival more likely. Barometric pressure declines predictably before inclement weather, and several lines of evidence indicate that animals alter behavior based on changes in ambient pressure. Here we examined the effects of declining barometric pressure on physiology and behavior in the white-crowned sparrow, Zonotrichia leucophrys. Using field data from a long-term study, we first evaluated the relationship between barometric pressure, storms and stress physiology in free-living white-crowned sparrows. We then manipulated barometric pressure experimentally in the laboratory and determined how it affects activity, food intake, metabolic rates and stress physiology. The field data showed declining barometric pressure in the 12–24h preceding snowstorms, but we found no relationship between barometric pressure and stress physiology. The laboratory study showed that declining barometric pressure stimulated food intake, but had no effect on metabolic rate or stress physiology. These data suggest that white-crowned sparrows can sense and respond to declining barometric pressure, and we propose that such an ability may be common in wild vertebrates, especially small ones for whom individual storms can be life-threatening events.

Key words: activity, corticosterone, environmental cues, inclement weather, metabolic rate, stress.

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INTRODUCTION

Organisms use environmental cues to time annual cycles, fine-tune breeding and make daily behavioral decisions. In the long term, predictable changes in photoperiod, rainfall and temperature help organisms correctly time life history transitions to maximize fitness. In the short term, organisms use a broader array of cues for predicting near-term weather. Here we focus on the latter class, asking whether sparrows use declining barometric pressure as a cue to prepare for oncoming storms.

Storms can profoundly affect foraging success, metabolic requirements, body condition and parental behavior. For example, heavy rains can flood nest sites and limit foraging opportunities, and high winds can decrease foraging efficiency and increase heat loss. Storms can also include low temperatures and snow. Cold temperatures can increase metabolic costs of thermoregulation and directly threaten survival. Snow can limit activity and cover food resources. Are there predictable components to these rapid abiotic changes? In principle, organisms could use storm-related cues over several time scales. The most proximate are derived from the storm itself – increasing cloud cover, high winds, falling temperatures and heavy precipitation. However, cues in advance of the storm may be more useful, because they potentially give organisms more time to prepare by appropriately altering physiology and behavior.

One well-known predictor of impending storms is falling barometric pressure. As inclement weather approaches, barometric pressure can decline by 2–12kPa over 24 to 72h [depending on the severity of the storm (Saucier, 2003)]. This phenomenon is well documented and could be a reliable cue used by vertebrates. Studies from a variety of taxa indicate that animals use barometric pressure to time behavioral transitions. Frogs may call more when barometric pressure is low and rain is likely (Brooke et al., 2000; Oseen and Wassersug, 2002); fish move into salt marshes (possibly to forage) as barometric pressure declines (Crinall and Hindell, 2004); and bats and birds can sense changes in barometric pressure (Kreithen and Keeton, 1974; Lehner and Dennis, 1971; Paige, 1995), and may alter migratory behavior to avoid poor weather (Blokpoel and Richardson, 1978; Cryan and Brown, 2007; Maransky et al., 1997; Matthews and Rodewald, 2010; Panuccio et al., 2010; Pyle et al., 1993; Sapir et al., 2011; Shamoun-Baranes et al., 2006). The majority of studies, however, examine correlations among behavioral patterns and naturally varying barometric pressure. Direct manipulation of barometric pressure itself is rare.

We also do not yet understand the mechanisms by which vertebrates integrate pressure information into physiological and behavioral responses. One likely candidate is the endocrine system, especially the adrenocortical axis, which is a well-known integrator of stress information in other contexts. Endocrine systems commonly...
translate environmental cues into organismal responses; for example, elevated androgens promote migratory behavior in response to longer days. Glucocorticoids are secreted from the adrenal gland, and are thought to redirect physiology and behavior so that animals can cope with deteriorating or unpredictable situations (Wingfield and Sapolsky, 2003). From a life history perspective, glucocorticoids are thought to redirect energy expenditures from reproduction to self-maintenance, increasing the likelihood of survival during suboptimal conditions (Breuner et al., 2008; Wingfield et al., 1998).

Several lines of evidence suggest that corticosterone (CORT; the primary avian glucocorticoid) regulates behavioral responses to storms. Animals captured during snow and rainstorms can have elevated CORT (Asheimer et al., 1995; Bize et al., 2010; Rogers et al., 1993; Smith et al., 1994; Wingfield et al., 1983). Extreme temperatures can increase circulating CORT (Bize et al., 2010; de Bruijn and Romero, 2011; Dunlap and Wingfield, 1995; Tyrrell and Cree, 1998). Glucocorticoids can increase activity in both laboratory and field settings, and have been shown to increase food intake in several vertebrates (e.g. Arvaniti et al., 1998; Asheimer et al., 1992; Breuner et al., 1998; Breuner and Hahn, 2003; Crespi et al., 2004; Nasir et al., 1999). And finally, glucocorticoid implants alter responses to inclement weather in white-crowned sparrows (Breuner and Hahn, 2003). Overall, these studies suggest that glucocorticoids could become elevated as barometric pressure changes and storms begin, and that rising glucocorticoids cause changes in physiology and behavior that enable animals to withstand deteriorating conditions.

Here we examine the relationships between barometric pressure, physiology and behavior in free-living and captive white-crowned sparrows. In a field study of wild sparrows, we used a 7-year data set to evaluate both how much barometric pressure declines before snowstorms, and the relationships between pressure and CORT physiology. In a laboratory study, we exposed birds experimentally to declining barometric pressure and evaluated changes in their metabolic rates, foraging behavior, activity and stress physiology.

**MATERIALS AND METHODS**

**Field study**

*Zonotrichia leucophrys oriantha* Oberholser 1932 were sampled as part of a long-term field study on stress physiology (Breuner and Hahn, 2003; Breuner et al., 2006; Crino et al., 2011; Hahn et al., 2004; Lynn et al., 2007). White-crowned sparrows are an ideal species to use for studies of environment–physiology–behavior interactions. Four of the subspecies (*gambelli, oriantha, leucophrys* and *pugetensis*) breed in habitats where spring storms are common, and individual birds often must redirect physiology and behavior to breed successfully (Addis et al., 2011; Breuner and Hahn, 2003; Romero, 2002; Wingfield et al., 1983; Wingfield and Ramenofsky, 2011). We have studied behavioral and physiological response to spring storms in *Z. l. oriantha* (the mountain white-crowned sparrow) since 1997.

Males *Z. l. oriantha* arrive at the high elevation breeding grounds in early May. Females arrive 2 weeks later and often begin laying eggs in early June (Morton, 2002). During this period, snow cover recedes from 100 to 50%, with frequent new snow. Birds were caught at Tioga Pass Meadow, Inyo National Forest, CA, USA (37°54′53″N, 119°15′18″W, ~3000 m elevation), with seed-baited potter traps. The majority of blood samples were collected between early May (when males first arrive at the breeding site) and mid-June (when nesting is underway and we no longer use seed-baited trap lines to catch birds). Blood was collected into heparinized capillary tubes from the alar vein after puncture with a 26 gauge needle, within 3 min of disturbance in the trap [sitting and eating in a potter trap does not alter baseline or <30 min CORT levels in white-crowned sparrows (Romero and Romero, 2002)]. Birds were held in a cloth bag, and serial blood samples were taken after 15 and 30 min [termed a ‘stress series’ (e.g. Wingfield, 1994)]. Blood was kept on ice until centrifuged (within 5 h), and plasma was removed and frozen until assayed. CORT levels were measured using enzyme immunoassay (EIA) as per Wada et al. (Wada et al., 2007). All assays were completed in the Breuner laboratory, using corticosterone EIA kits from Assay Designs (ADI-901-097, Enzo Life Sciences, Farmingdale, NY, USA). The data presented here are collated from over 8 years of assays; on average detectability levels were between 0.5 and 1.5 ng ml⁻¹, average intra-assay variation was 6.6%.

**Laboratory experiment**

*Z. l. oriantha* (the mountain white-crowned sparrow) since 1997. If sparrows respond to approaching storms by eating more, they may also gain mass or fat. However, it is also possible that the declining environmental conditions would require greater energy expenditure to maintain body temperature. We evaluated the direction of the relationship between barometric pressure change and mass or fat scores using linear mixed-effects (Lme) models (Pinheiro and Bates, 2000), with days before the first egg lay of the season as a covariate, and individual ID as a random factor (R2.11.1, package nlme).

**Barometric pressure versus mass and fat scores**

To determine the relationship between barometric pressure and stress physiology in free-living birds, we calculated the change in barometric pressure over the 12 h prior to each stress series sampled. Both baseline CORT (endogenous level of CORT measured on capture, N=773), and maximum CORT (highest CORT measured in response to handling stress, N=452) were compared with the rate of barometric pressure decline during the preceding 12 h. This analysis evaluates barometric pressure effects on both (1) resting levels of CORT and (2) the animals’ ability to respond to subsequent stressors.

**Storms**

To evaluate the barometric pressure change prior to snowfall, we identified six storms from 2002 to 2008 when first snowfall was detected at the meadow. We then graphed barometric pressure from the previous 12+ h.

**Barometric pressure measured in Tuolumne Meadows, Yosemite National Park (37.9°N, 119.4°W), by Dr Jessica Lundquist (Department of Civil and Environmental Engineering, University of Washington) as part of a long-term environmental study [~1 km from Tioga Pass Meadows (Lowry et al., 2010)]. Barometric pressure was logged every 30 min from 2001 to present.

**Storms**

To determine the relationship between barometric pressure and stress physiology in free-living birds, we calculated the change in barometric pressure over the 12 h prior to each stress series sampled. Both baseline CORT (endogenous level of CORT measured on capture, N=773), and maximum CORT (highest CORT measured in response to handling stress, N=452) were compared with the rate of barometric pressure decline during the preceding 12 h. This analysis evaluates barometric pressure effects on both (1) resting levels of CORT and (2) the animals’ ability to respond to subsequent stressors.
Chambers and pressure manipulation

Birds were put into custom-built Plexiglas chambers (32×30.5×32 cm outside, 30.5×29×30.5 cm inside; University of Texas machine shop, Austin, TX, USA) fitted with several doors (for entry into the cage and for food and water) and several ports for gas flow and pressure measurements. All doors were sealed with O-rings and ports with airtight bulkhead fittings.

The experimental setup is shown in Fig. 1A. Chambers were ventilated by ~3 m³ min⁻¹ CO₂-free air, which was pulled by a vacuum system and controlled by an upstream solenoid valve. CO₂-free air was produced by a gas generator (PureGas, Broomfield, CO, USA) at 3.5–41 m³ min⁻¹ and pushed to an overflow bypass. The vacuum pulled air from the overflow through the bird chamber and then through a Venturi flow tube, which we used as a flow meter. The Venturi tube consisted of two copper tubes soldered together with a short, conical copper connector; the upstream tube was larger (7.2 mm diameter) than the downstream tube (4.5 mm). Each section had a port drilled into it to which short pieces of small brass tubing were fitted. These two lines were then connected to a differential pressure meter (PT-100B, Sable Systems, Las Vegas, NV, USA).

The arrangement was calibrated by simultaneously measuring flow rate of air (with a calibrated fixed rotameter, max. 100 ml min⁻¹) and differential pressure in the Venturi tube (with the differential pressure meter). As expected, the relationship between flow and differential pressure was exponential (Fig. 1B); to linearize, we log transformed both sides (Fig. 1B, inset). The relationship was highly repeatable across days (the calibration includes measurements from multiple days). Once the calibration curve was obtained, we used only differential pressure to estimate flow rates, as this was more easily logged onto a computer.

We manipulated pressure in the bird chamber by controlling the degree of opening of a solenoid valve placed upstream. A vacuum pulled air through the system at an approximately constant rate; therefore, greater resistance upstream led to a greater drop in pressure from room to chamber. The solenoid valve was under computer control, and we therefore could automatically ramp down the pressure in the chambers to mimic an approaching storm (Fig. 1C). Absolute pressure in the chambers was monitored continuously (PT-1000, Sable Systems).

Metabolic measurements

We estimated metabolic rates from rates of CO₂ emission. Outgoing air from the Venturi air tube was sub-sampled at 200 ml min⁻¹ (MFS-1, Sable Systems). To remove water vapor, the sub-sampled stream was pushed through a tube of Drierite. CO₂ content of the stream was then measured by an infrared gas analyzer (CA-2A, Sable Systems), with data logged at 12 Hz onto a computer via an analog-digital converter (UI-2, Sable Systems). The gas analyzer was calibrated frequently (zero gas was pure N₂ and span gas was 2800 p.p.m. CO₂ in N₂).

Because each bird was kept for several days in its Plexiglas chamber, baselining was performed in two stages. Before and after each run, the analyzer was baselined with pure N₂ (to measure drift in the analyzer). To obtain baseline levels of CO₂ leaking into the rest of the system (chamber + other parts), chambers were sealed as in an experiment, but without birds, and then ramped through typical pressure changes. Leaks were large enough that chambers had background levels of 50–60 p.p.m. CO₂. However, the levels were not significantly affected by pressure ramping. Moreover, levels of CO₂ from chambers containing birds were much higher, from 600 to 1300 p.p.m. To adjust metabolic traces from each bird, we subtracted its chamber background value from measured experimental values. These baseline-adjusted values were then converted to rates of CO₂ emission by multiplying the molar fraction of CO₂ by the molar flow rate. We did an additional transformation to account for the large chamber volume (28.3 l) compared with the
overall flow rate (~31 min⁻¹), which has the effect of integrating metabolic events over long periods of time (the half-life of CO₂ in the chamber was 9.4 min=28.3 min⁻¹/3 min⁻¹). To obtain instantaneous rates of CO₂ emission, we used eqn3 from Bartholomew et al. (Bartholomew et al., 1981), with effective chamber volume set to 28.31 and the other parameters (actual flow rate, CO₂ levels) measured during the experiments. The equation calculates future equilibrium levels of CO₂ from the short-term kinetics of change at any instant.

Experimental protocol
Birds were housed in the Plexiglas housing chamber 3 days prior to running the experiment to allow acclimation to the new housing. The testing room held two Plexiglas chambers, each holding one bird; birds were visually isolated from each other during this time. Only one bird was tested each day, and each bird was tested two days in a row (control and pressure change, order of treatment randomized across birds). Birds were videoed from outside the testing room, through a one-way mirror. On testing day, the video was started at ~10:00h. One hour later, the computer program was activated to begin barometric pressure decline. After 3 h of barometric pressure decline, the video was stopped, blood samples were collected and birds were returned to the housing chamber. On control testing days the exact protocol was followed, but barometric pressure levels were left at ambient (see Fig. 1D).

Behavior
Videos were scored for five behaviors: (1) activity: the number of times the bird crossed the vertical midline of the cage; (2) foraging: the number of pecks made into the covered food dish; this was not precisely food intake, as ingestion of food was not ascertained with each peck; (3) water intake: the number of drinks taken from the water dish; (4) bill swipe: each set of two swipes (brushing the bill against the perch or other object, once left to right and once right to left) was counted as one bill swipe; and (5) random peck: any peck to a surface other than inside the covered food dish.

We also analyzed videos using an automated technique, which provided higher temporal resolution (1 s) at the cost of less behavioral detail. Raw digital videos (30 frames s⁻¹) were processed in a VLC media player (v. 2.0.4, http://www.videolan.org/vlc/) to extract 1 frames s⁻¹. The image sequences were then imported into ImageJ (v. 1.46r, National Institutes of Health, Bethesda, MD, USA) and processed, using a set of differenting operations, to produce a video containing only white and black pixels, with white corresponding to the difference between successive images (see ImageJ script in supplementary material Appendix S1). The mean pixel value of each frame was then outputted to a text file for further processing in R. This analysis provided detailed information about overall movements of birds within each single run, but could not identify particular movements (as was done by eye). Examples of raw and processed videos and a plot of the outputted behavioral data are available in supplementary material Movies 1, 2 and Fig. S1).

CORT sampling
At the end of the 4 h, we obtained a blood sample from the focal bird within 3 min of entering the room. Birds were then held in a cloth bag, and serial blood samples were taken 15 and 30 min after initial sampling. Blood was immediately placed on ice until it was centrifuged (within 1–2 h). Plasma was removed and assayed with EIA as described above for the field samples.

Statistical analysis
Field data (barometric pressure change and CORT) were analyzed using lme models (Pinheiro and Bates, 2000) (R2.11.1, package nlme) with individual as a random factor. CORT values from the field study were In transformed, and statistical outliers (mean ±2 s.d., N=32 of 805 total data points) were removed from the baseline CORT data. Inclusion of outliers did not change the reported relationship. CORT values in the laboratory study were log+1 transformed to account for heteroscedasticity and then analyzed with lme, with individual as a random factor.

Metabolic data also were analyzed using lme models implemented in R, with hour and treatment (control or pressure) as main effects and individual as a random factor. In this analysis, the key statistic of interest is whether there is a significant interaction between time and treatment, which (if present) could indicate that the metabolic rates of birds in the two treatments change in different ways over time. We followed a standard protocol for model identification (Crawley, 2007). In brief, metabolic rate was modeled as a function of the main effects of activity, treatment and hour, with individual specified as a random effect. The metabolic data were extensively autorecorrelated, reflecting that metabolic rate at a given time more closely resembled other metabolic rates nearby in time compared with metabolic rates at more distant times. An initial model not accounting for autocorrelation had residuals that were significantly autorecorrelated up to lags of many minutes. A subsequent set of mixed-effects models were specified with different autocorrelation structures (including autoregressive moving averages of orders 2–5, and autoregressive of order 1), and those models were compared using the Akaikes’s information criterion (AIC). Once the best model was identified (lowest AIC), it was validated by examining residual structure. The total size of the data set (~32,000 observations) required memory allocation far in excess of what was available to us; we therefore subsampled every 10th data point.

Behavioral data were analyzed in two main ways. Visually scored data were binned into hours for each bird. Data were then corrected for day-to-day variation in individual behavior (e.g. food pecks in hour 1 subtracted from food pecks in hours 2, 3 or 4 on that day), so that elevations or declines in behavior over baseline levels were analyzed. Baseline-corrected values were compared across treatments using paired t-tests (GraphPad Prism, GraphPad Software, La Jolla, CA, USA). Effect size (Hedges’ d) was calculated using MetaWin 2.1 http://www.metawinsoft.com/ (Hedges and Olkin, 1985). All data were normally distributed, except for activity, which was log transformed before analysis.

The pixel-based data were analyzed using measures of cumulative activity, considered between treatments within birds. The null expectation is that cumulative activity increases linearly with the same slope between treatments. If pressure change leads to changes in activity levels, this will be reflected as a shift in the slope toward the end of the curve in the pressure treatment (e.g. if activity increases during pressure change, the cumulative activity will curve upward). A secondary consideration, which emerged from visual inspection of the behavioral data, was whether bouts of activity were periodic; we analyzed this, for all behavioral traces, using autocorrelation functions.

Finally, to examine whether activity influenced metabolic rate, we used cross-correlation analysis between the time series of metabolic rates and their associated time series of high-resolution (pixel-based) behavior. The behavioral data were first averaged into 5s bins (so that their frequency would match that of the metabolic data obtained at 12Hz) and then pre-whitened by differencing once, which removed small-amplitude shifts in baseline but kept the spikes.
RESULTS

Field data

Barometric pressure and storms

During the 12 h prior to snowfall, barometric pressure declined on average by 2.31±1.02 kPa (range 2.88 to 1.34, N=6; Fig. 2). The slope (pressure change per hour) for each storm was significantly different from zero (linear regression, P<0.0001 in each case, average F=449±129).

There was no relationship between barometric pressure change over 12 h and endogenous CORT secretion in free-living white-crowned sparrows (lme model with individual as a random effect; time: F1,409=0.77, P=0.79; maximal CORT: F1,409=0.702, P=0.403; Fig. 3).

There was no effect of barometric pressure decline on mass in the free-living population (lme: F1,362=0.06, P=0.8). However, there was a relationship between fat score and barometric pressure change (lme: F1,370=15.26, P<0.0001, significant interaction with days before first egg lay: F1,370=96.07, P<0.0001). The relationship was positive, in that birds sampled when the barometric pressure has been declining had lower fat scores; however, the r² was very low (0.014).

Laboratory experiment

Metabolic rate

We measured metabolic rates continuously during 4 h trials in the first year (N=6). Metabolic rate during each run (second, third and fourth hours) did not change from baseline (first hour) rates in either the control or barometric pressure change treatments (Fig. 4A,B, supplementary material Figs S2, S3). These data were analyzed using lme models, with individual as the random factor. The key test was whether there was a significant interaction between hour and treatment, which would indicate that the metabolic rates in one treatment group changed (over time) differently than those in the other treatment group. In fact, that term was insignificant (Table 1), so we concluded that there was no effect of pressure on metabolic rate. The significant effect of treatment does not reflect real differences arising as a cause of the treatments; rather, it reflects that by chance the birds had statistically different metabolic rates during the first hour (before the treatments commenced). With no effect of pressure evident in the first year, we decided against measuring metabolic rates in the second year.

Stress physiology

Blood samples were taken at 0–3, 15 and 30 min after the experiment ended. There was a significant increase in CORT over the 30 min, but no effect of treatment, and no interaction between time and treatment (lme with individual as a random factor; time: F1,65=495, P<0.0001; treatment: F1,65=1.49, P=0.23; interaction: F2,65=0.33, P=0.72; Fig. 5). Integrated CORT (the entire amount of CORT secreted over the 30 min) also did not vary by treatment (paired t-test; t13=1.20, P=0.25; Fig. 5, inset).

Behavior

Visually scored behavioral results (sum of counts per hour, corrected for baseline behavior that day) are shown in Table 2. A positive number indicates that behavior increased from hour 1, a negative number indicates a decrease. There was no significant effect of treatment on activity, random pecks (not to food), water intake or bill swipe (P>0.25, P=0.14, P=0.27 and P>0.21, respectively).

Foraging increased significantly during the third hour of pressure treatment (t13=2.611, P=0.02; Fig. 6B), but levels were not significantly different during hours 2 or 4 (t13=0.51, P=0.62 and t13=0.94, P=0.36, respectively).

The high-resolution (1 Hz) video analyses showed no effect of treatment on activity. Most of the cumulative activity traces (supplementary material Fig. S4) rose approximately linearly, indicating that rate of large movements did not change with time. Two of six birds had cumulative activity traces whose slopes became shallower during the pressure change, indicating reduced movement late in the session. One of the six birds had a cumulative activity trace that became shallower as the session progressed during the control treatment.

The pixel-based analyses also suggested that bird movements were periodic (see Fig. 4A,B, supplementary material Figs S2, S3). We analyzed this periodicity quantitatively using fast Fourier transforms implemented in R. All periodograms had the shape of the one shown in Fig. 4C, with most of the power concentrated at low frequencies (corresponding to long periods between bouts of...
in all periodograms, we picked out the single frequency containing the highest power, for each bird × treatment combination, and converted those frequencies to periods in minutes. The mean value was 31.52 min (range 17.35–48.00), and there was no difference according to pressure treatment.

Correlations between activity and metabolic rate
Although neither metabolic rate nor activity changed significantly as functions of time or treatment, inspection of the traces together (Fig. 4A,B, supplementary material Figs S2, S3) suggested that the two measures were strongly correlated. Cross-correlation analysis of the time series confirmed this impression (example in Fig. 4D). Of the 12 pairs of time series (two pairs per bird × six birds), nine showed major peaks of cross-correlation centered around lags of zero. Of the remaining three that did not show strong cross-correlation, two were pressure treatments and one was a control. The larger bouts of activity raised instantaneous rates of CO2 emission by 35–100% (see supplementary material Figs S2, S3).

DISCUSSION
Declining barometric pressure reliably signals approaching storms. We examined whether, in anticipation, sparrows altered their behavior or physiology. We predicted that glucocorticoid hormones (CORT in birds) would respond to this environmental change, possibly mediating the behavioral and physiological reactions. In white-crowned sparrows, declining barometric pressure indeed altered behavior. Compared with control birds, those in the pressure treatment increased foraging behavior. However, those birds showed no changes in metabolic rate or CORT physiology.

Our data thus provide modest support for the hypothesis that animals can use changing barometric pressure to predict the onset of inclement weather. Several studies indicate that birds can detect changes in barometric pressure (Kreithen and Keeton, 1974; Lehner and Dennis, 1971), and may alter behavioral patterns as a result

**Table 1. Summary of linear mixed-effects model of the metabolic data shown in Fig. 4A,B and in supplementary material Figs S2, S3 (all metabolic data)**

<table>
<thead>
<tr>
<th>Source</th>
<th>( F_{1,3272} )</th>
<th>( P )</th>
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</thead>
<tbody>
<tr>
<td>Intercept</td>
<td>3514</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Treatment</td>
<td>33.4</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Hour</td>
<td>0.84</td>
<td>0.43</td>
</tr>
<tr>
<td>Hour × Treatment</td>
<td>0.13</td>
<td>0.71</td>
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</table>

**Fig. 5. Corticosterone (CORT) levels 0–3, 15 and 30 min after completion of the barometric pressure experiment. Data shown are means ± s.e.m. Inset: integrated CORT over the entire 30 min. Birds were held in cloth bags between sampling times.**
Foraging energy at a higher rate as conditions decline. However, the decline in the wild. This indicates that sparrows may be spending more or have greater fat stores. Our data indicate that although free-living birds caught after pressure decline in the field may increased food intake in our laboratory experiment, we may expect energy stores before the storm prevents further foraging. With unpredictable or stressful events (Breuner et al., 2008; Wingfield et al., 2006). However, homing requires a map and a compass [vision and magnetic detection (Kramer, 1953)], but not a barometer. Several other studies have hypothesized that the PTO detects barometric pressure (Paige, 1995; Shamoun-Baranes et al., 2006; von Bartheld and Giannessi, 2011), but none have tested it directly.

CORT physiology did not correlate with barometric pressure change in the field, and experimental barometric pressure decline in the laboratory did not affect circulating or inducible glucocorticoids. This is surprising, given the role glucocorticoids play in shifting time and energy allocation towards survival during unpredictable or stressful events (Breuner et al., 2008; Wingfield et al., 1998; Wingfield and Sapolsky, 2003). However, there was clearly no relationship between barometric pressure and CORT in free-living sparrows, and CORT levels were highly overlapping in the laboratory experiment.

Metcalfe et al. (J. Metcalfe, K. L. Schmidt, W. Bezner Kerr, C. G. Guglielmo and S. A. MacDougall-Shackleton, submitted) have also recently tested the behavioral and physiological responses to barometric pressure change in laboratory sparrows. Their experimental design differed substantially from ours, but the results were similar. In white-throated sparrows (Zonotrichia albicollis, a sister species to the white-crowned sparrow), a rapid decline in barometric pressure decreased the latency to feed, but had no effect on glucocorticoid physiology.

Few studies have evaluated correlations between environmental barometric pressure and endogenous glucocorticoids. Frigerio et al. (Frigerio et al., 2004) discovered a significant relationship between barometric pressure and CORT metabolites appearing in the feces of graylag goose (Anser anser). Given the arguments presented here, we would expect that low barometric pressure is associated with elevated CORT. However, Frigerio et al. found the opposite effect: low barometric pressure during the preceding morning was

<table>
<thead>
<tr>
<th>Behavior</th>
<th>Second hour</th>
<th>Third hour</th>
<th>Fourth hour</th>
</tr>
</thead>
<tbody>
<tr>
<td>Activity</td>
<td>–30.8±32.9</td>
<td>–39.4±24.7</td>
<td>–23.9±33.4</td>
</tr>
<tr>
<td>Pressure decline</td>
<td>–23.0±13.4</td>
<td>–1.14±23.7</td>
<td>–41.0±15.4</td>
</tr>
<tr>
<td>Water intake</td>
<td>–0.9±16.9</td>
<td>–12.5±16.2</td>
<td>–40.7±36.5</td>
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<td>Random peck</td>
<td>–0.14±0.98</td>
<td>–0.71±0.91</td>
<td>–0.65±0.64</td>
</tr>
<tr>
<td>Pressure decline</td>
<td>0.62±0.90</td>
<td>–0.86±0.77</td>
<td>–0.50±0.82</td>
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<td>Bill swipe</td>
<td>11.0±26.0</td>
<td>–1.87±4.34</td>
<td>9.50±14.2</td>
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<td>Pressure decline</td>
<td>–10.2±17.4</td>
<td>–25.4±12.8</td>
<td>–17.2±16.5</td>
</tr>
<tr>
<td>Corrected no. of food pecks (third hour)</td>
<td>–18.2±13.0</td>
<td>–14.1±9.21</td>
<td>–11.0±6.83</td>
</tr>
</tbody>
</table>

Data are means ± s.e.m. of the sum of behaviors over the hour, corrected for baseline (N=13 for second hour, N=14 for third and fourth hours).

P<0.05 with paired t-test.

Foraging behavior was elevated during the third hour of declining barometric pressure as compared with the control treatment. (A) Absolute number of food pecks during the third hour. (B) The same data corrected for hour 1 behavior.
associated with low CORT metabolites in the feces the next morning. The authors hypothesized that low barometric pressure promoted a ‘sit and wait’ strategy, which in turn resulted in lower levels of CORT (activity promotes CORT secretion). It is also possible that barometric pressure decline increased food intake, thereby diluting the concentration of CORT in the feces. This is the only field CORT–barometric pressure paper that we have identified. Romero et al. (Romero et al., 2000) examined correlations between CORT secretion and several weather variables in three species of arctic passerine, but did not consider barometric pressure.

In summary, our data demonstrate that white-crowned sparrows respond to declining barometric pressure by altering their behavior in ways that appear to prepare them for inclement weather. We found no association between barometric pressure and CORT-driven stress physiology in either the field or the laboratory. It is possible that other neuro-hormonal systems [e.g. orexin, neuropeptide Y, agouti-related peptide (e.g. Thorpe and Kotz, 2005) (for a review, see Arora and Anubhuti, 2006)] drive this system. A more neurological approach in ways that appear to prepare them for inclement weather. We found no association between barometric pressure and CORT-driven stress physiology in either the field or the laboratory. It is possible that other neuro-hormonal systems [e.g. orexin, neuropeptide Y, agouti-related peptide (e.g. Thorpe and Kotz, 2005) (for a review, see Arora and Anubhuti, 2006)] drive this system. A more neurological approach.


**Fig. S1.** Mean image pixel value over the course of Movie 2. The script generating these data is given in Appendix S1. Greater pixel values mean that more pixels differed between one frame and the next, indicating larger movements by the bird.
**Fig. S2.** Rates of CO₂ emission for birds in their control (blue) or pressure (red) treatment. Corresponding behavioral traces, from the pixel-based method of video analysis, are plotted below each metabolic trace. Data from the second set of three birds is shown in Fig. S3.
Fig. S3. Continuation of the data shown in Fig. S2.
Fig. S4. Cumulative activity plots from the pixel-based video method. Each bird’s data is shown in a single plot, in both blue (control) and red (pressure).
**Movie 1.** Video of 400 s of a white-crowned sparrow. Frames were extracted (1 frame s$^{-1}$) from the original 30 frames s$^{-1}$ digital video, and they are played back at 8 frames s$^{-1}$ here (8× compression of time).

**Movie 2.** Video of the same 400 s but after the file (Movie 1) was processed in ImageJ using the script shown in Appendix S1. Each time the bird moves, the difference between its position in the current versus the prior frames shows up as white pixels. The script also calculates the mean pixel value of the entire frame and outputs it to a text file. The data for this video segment is plotted in Fig. S1.
Appendix S1. ImageJ script used to process videos.

```java
function processbird(path){
    rename("A");
    stacksize = nSlices;
    run("Z Project...", "start=1 stop=&stacksize projection=[Max Intensity]");
    rename("Z");
    imageCalculator("Difference create stack", "A", "Z");
    rename("B");
    imageCalculator("Copy create stack", "B", "B");
    rename("C");
    selectWindow("B");
    run("Delete Slice");
    selectWindow("C");
    setSlice(stacksize);
    run("Delete Slice");
    imageCalculator("Difference create stack", "B", "C");
    run("Add to Manager");
    makeRectangle(3, 2, 357, 238);
    roiManager("Add");
    roiManager("Multi Measure");
    saveAs("Results", path+"results.txt");
    if (isOpen("Results")) {
        selectWindow("Results");
        run("Clear Results");
        run("Close");
    }
    selectWindow("ROI Manager");
    run("Close");
    selectWindow("A");
    run("Close");
    selectWindow("B");
    run("Close");
    selectWindow("Result of B");
    run("Close");
    selectWindow("C");
    run("Close");
    selectWindow("Z");
    run("Close");
}
sc=50; //this switch sets image size to 50%, which saves a lot of memory
path="C:/Users/john.doe/Desktop/yourimages/";
run("Image Sequence...", "open=&path scale=&sc sort");
processbird(path);
```

Appendix S1. ImageJ script used to process videos.